

SHORT COMMUNICATION

TRITERPENES FROM MOSSES—I.

THE OCCURRENCE OF 22(29)-HOPENE IN *THAMNIUM ALOPECURUM*
(L.) BR. EUR. SSP. *EU-ALOPECURUM* GIAC.

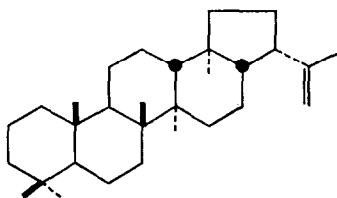
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To our knowledge only few triterpenes have so far been isolated from mosses.¹ We wish to report here a preliminary chemical examination of the moss *Thamnium Alopecurum* (L.) Br. eur., ssp. *eu-Alopecurum* Giac., as a part of a wider chemotaxonomic examination of the most common Bryophyta of the Italian flora.

The unsaponifiable portion of the light petroleum extract of the plant was chromatographed over alumina; light petroleum eluted 22(29)-hopene (I). Elution with ethyl ether



(I)

afforded four fractions containing mixtures of alcohols; ergosterol, stigmasterol and β -sitosterol were identified in all fractions by GLC retention times and i.r. spectra. At least two other unidentified alcohols (triterpenes, or more probably sterols) were also present; these are still under study.

The fatty acids obtained from the saponified material were converted into the corresponding methyl esters and analysed by GLC: the presence of acids from C₁₂ to C₂₈ (some of which unsaturated) was established.

Triterpene hydrocarbons have so far been isolated only from ferns² and, in one instance, from a lichen.³ The occurrence of 22(29)-hopene in *Thamnium* is of interest, as this is the first example of a triterpene hydrocarbon isolated from a moss. A more extensive survey would be desirable, to test the hypothesis that these compounds are characteristic of primitive plants.

¹ R. HEGNAUER, *Chemotaxonomie der Pflanzen*, p. 188. Birkhauser Verlag, Basel (1962).

² H. AGETA, K. IWATA and S. NATORI, *Tetrahedron Letters*, 1447 (1963); H. AGETA, K. IWATA and K. YONEZAWA, *Chem. Pharm. Bull. (Tokyo)* **11**, 408 (1963); H. AGETA, K. IWATA and S. NATORI, *Tetrahedron Letters*, 3413 (1964); G. BERTI, F. BOTTARI, B. MACCHIA, A. MARSILI, G. OURISSON and H. PIOTROWSKA, *Bull. Soc. Chim. Fr.* 2359 (1964); G. BERTI, F. BOTTARI, A. MARSILI, I. MORELLI and A. MANDELBAUM, *Chem. Commun.* 50 (1967); G. N. PANDEY and C. R. MITRA, *Tetrahedron Letters*, 4683 (1967).

³ T. BRUUN, *Acta Chem. Scand.* **8**, 1291 (1954).

EXPERIMENTAL

The dried plant (1 kg), collected in winter, was extracted in a soxhlet apparatus with light petroleum (6 l., b.p. 30–60°) for 40 hr. Concentration of the extract to a small volume led to separation of a wax (0.2 g, λ_{CO} 5.79 μ). Evaporation to dryness gave a brown oil (3.6 g) which was boiled under reflux with 10 per cent NaOH–EtOH (30 ml) for 4 hr. The alkaline soln. was diluted with H₂O and extracted with Et₂O. The extract, on evaporation, afforded 1.2 g unsaponifiable material. From the alkaline soln., by acidification with 2 N H₂SO₄ and new extraction with Et₂O, 1.1 g fatty acids were obtained.

Unsaponifiable Fraction

The unsaponifiable material was dissolved in light petroleum (100 ml, b.p. 30–60°) and chromatographed on neutral alumina (105 g, grade I, column 1.5 × 53 cm). By elution with light petroleum (500 ml) and evaporation of the solvent, a semi-solid residue was obtained. This was crystallized from acetone and from CHCl₃–MeOH to afford needles (25 mg), m.p. 211–214°, $[\alpha]_D^{20} + 59.5^\circ$. I.r., $\lambda_{>C=CH_2}$ 6.10, 11.26 μ , NMR, CH₂=C(CH₃)— 81.73 (3H, m), 4.75 (2H, m) ppm. The product was identified as 22(29)-hopene (hopene-b) by comparison with an authentic sample.⁴

Benzene did not elute any product; Et₂O was then used as eluent. From the first 300 ml, 0.2 g semi-solid residue (A) was obtained. Then, three fractions of 80 ml each were collected; these contained, respectively, 0.12 (B), 0.18 (C) and 0.09 g (D) residue. Further elution with Et₂O and with Et₂O–MeOH (9:1 v/v) did not yield any other product. The residues A–D contained mixtures of alcohols (i.r.). These were converted into the corresponding trimethylsilyl ethers and gas-chromatographed with a Perkin–Elmer F 20 instrument (column, 3 per cent SE 30 silicone gum rubber,⁵ temp. 250°, carrier gas N₂, flow rate 25 ml/min). Approximate compositions (per cent):⁶ A, ergosterol (10), stigmasterol (6), β -sitosterol (66), unknown alcohol I (12), unknown alcohol II (5); B, ergosterol (33), stigmasterol (9), β -sitosterol (57); C, ergosterol (39), stigmasterol (19), β -sitosterol (41); D, ergosterol (38), stigmasterol (28), β -sitosterol (33). These mixtures were also identified in part by comparison of their i.r. spectra with those of authentic samples. The i.r. spectrum of fraction A showed bands at 6.10 and 11.25 μ , typical of a $>C=CH_2$ group.

Fatty Acids

The fatty acids were esterified with CH₂N₂ in Et₂O, and the mixture of methyl esters was gas-chromatographed with a Perkin–Elmer 800 instrument (column, 20 per cent ethylene glycol succinate on acid-washed Chromosorb W, temp. 210°, carrier gas He, flow rate 40 ml/min). The following acids were identified (per cent): lauric (1.2), myristic (0.6), palmitic (29.0), palmitoleic (2.0), stearic (1.8), oleic (16.0), linoleic (22.0), linolenic (6.0), arachidic (1.2), behenic (1.8), erucic (9.5), lignoceric (3.0), cerotic (2.5), montanic (0.9).

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⁴ W. J. DUNSTAN, H. FAZAKERLEY, T. G. HALSALL and E. R. H. JONES, *Croat. Chem. Acta* **29**, 173 (1957).

⁵ This column proved to be the most suitable for a good separation of the sterols. Inferior results were obtained by using a neopentyl glycol succinate on Chromosorb W column; anyway, the presence of ergosterol, β -sitosterol and stigmasterol was confirmed in all fractions.

Retention times of known products were compared with those of authentic samples.